

Development of a Bioluminescent Cell-based Bioassay to Measure Fc Effector Function in ADCC

Terry Surowy, PhD
Research Manager, Promega Corporation

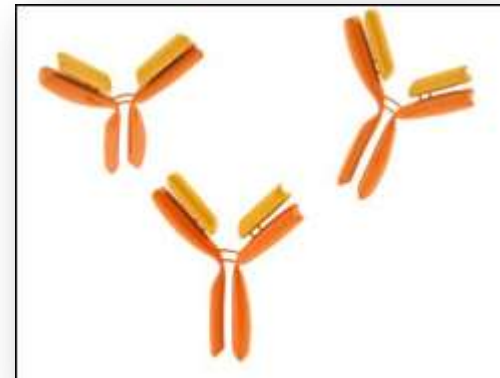


A new bioassay for Fc effector functionality in ADCC



Highlights of a novel bioluminescent cell-based bioassay:

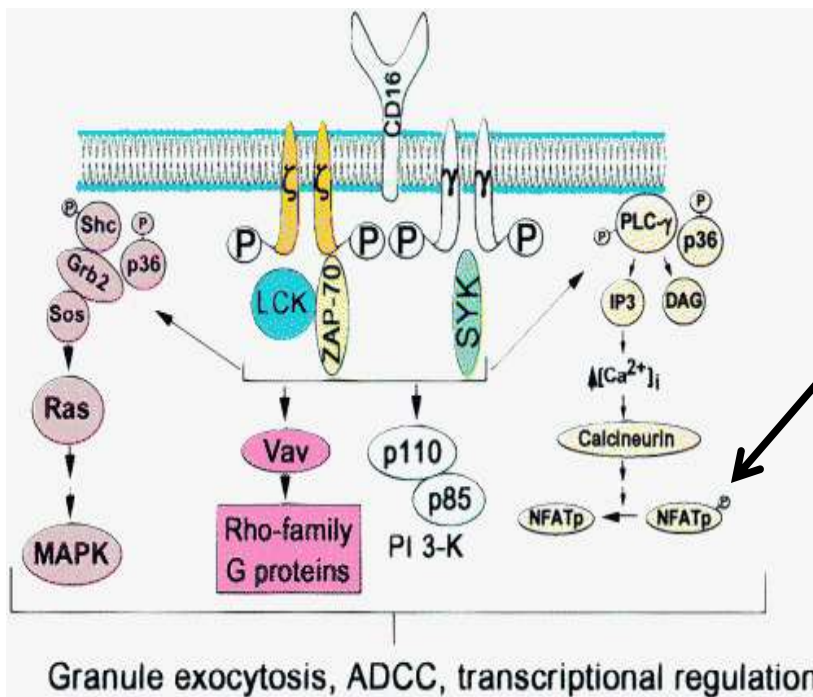
- A cell-based bioassay to quantify potency of Fc effector function in ADCC MOA pathway
- Based on target cell-bound antibody activation of FcγIIIa receptor signaling pathways in the effector cell
- Bioluminescent NFAT-RE-luciferase reporter bioassay readout is in engineered effector cells



- The bioassay is robust, specific, precise and accurate
- Suitable for stability studies, lot release and antibody characterization
- Effector cells are reagents -in frozen, thaw-and-use format

Signaling pathways activated in effector cells when FcγRIIIa is bound by target cell-bound antibody in ADCC

FcγRIII initiated signaling events



Leibson-PJ, *Immunity* 1997

New reporter gene bioassay uses an earlier step in the pathway

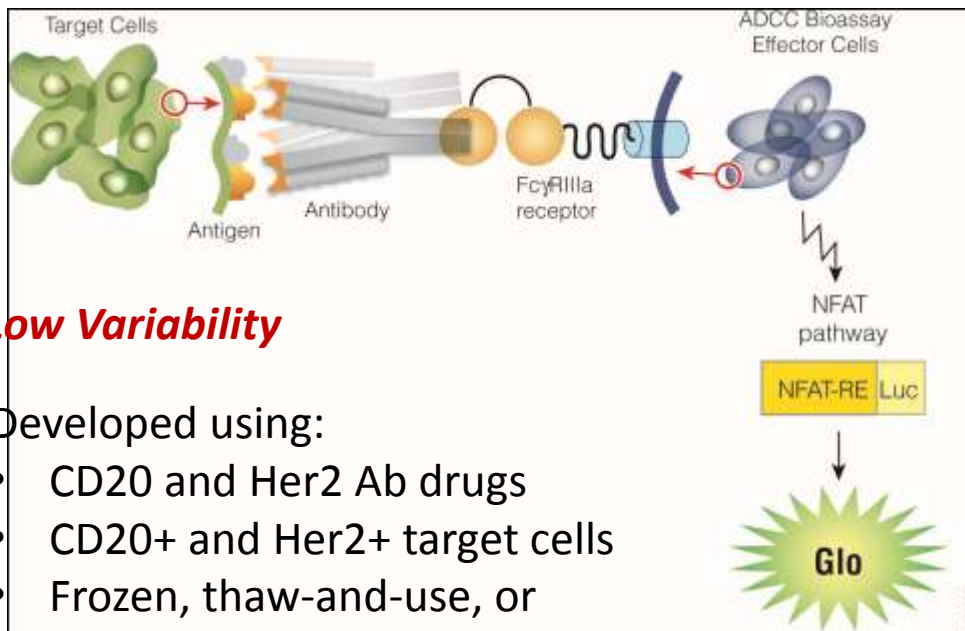
- To quantify biological activity of Abs in this MOA pathway
- Avoids preparation and high variability of primary effector cells
- Avoids high background and variability of spontaneous lysis

Traditional readout is cytotoxicity, the ultimate endpoint

Introduction to the reporter bioassay for Fc effector function in MOA pathway



NFAT-RE luciferase bioassay



Low Variability

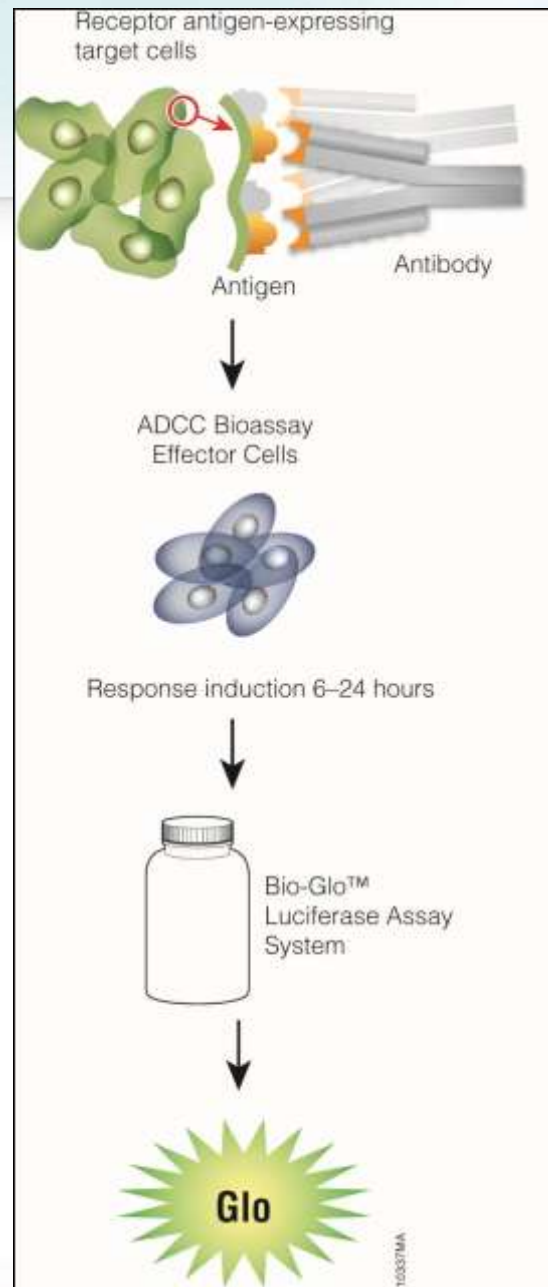
Developed using:

- CD20 and Her2 Ab drugs
 - CD20+ and Her2+ target cells
 - Frozen, thaw-and-use, or continuously cultured cells
-
- Extensive 'alpha' evaluations:
 - tested in multiple global biopharma & biotechs
 - tested in multiple systems

1. Effector cells are engineered to express FcγRIIIa (V158) and NFAT-RE-luc2 luciferase
2. 'Cells as reagents' (thaw-and-use)
3. Homogeneous assay format – simple 'add-mix-read' bioluminescent assay.
4. Optimized and robust assay reagents and protocol
5. Performance characteristics that meet needs of stability testing, lot release and Ab characterization

Protocol

1. Reference or test antibody is added to target cells.
2. Effector cells are added and response is induced in as little as 6 hours. Effector cells consist of a Jurkat cell line engineered to express FcγRIIIa (V158) receptor and NFAT-RE luc2 luciferase.
3. Luciferase detection reagent is added and luminescence is measured immediately.



Cells as reagents in bioassays

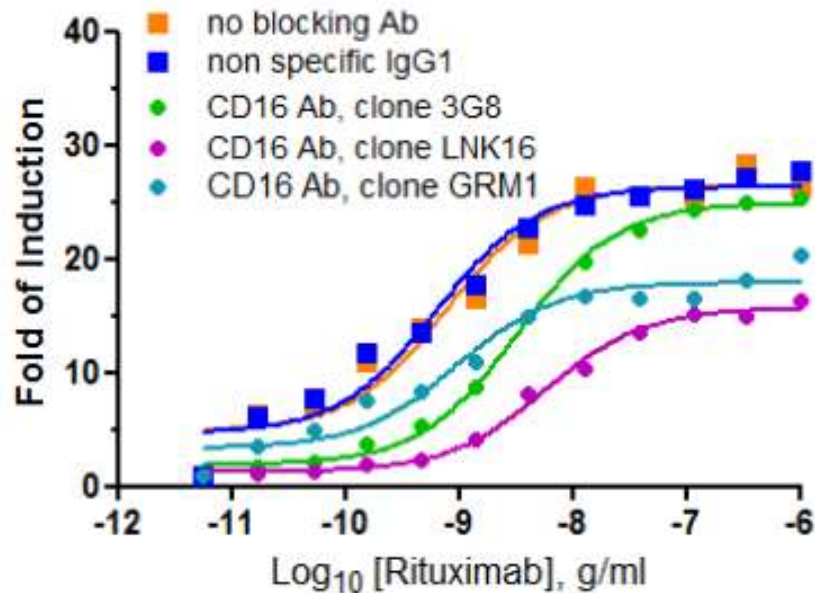
Frozen, Thaw-and-Use Cells

1. Human cell lines
 - Developed as Thaw-and-Use for immediate use in bioassay
 - Designed to give good recovery and robust response upon thawing
2. Thaw-and-Use format
 - Cell propagation conditions & defined freezing protocol control assay performance for a consistent bioassay response
 - No pre-culturing prior to assay means less variability introduced (overgrown, etc.)
 - Indefinite storage
 - Identical cells in bioassay, day-to-day
3. Minimizes pre-assay planning, time & labor
 - Ample cell banks provide long-term supply

FcγRIIIa antibody blocking of ADCC reporter bioassay response demonstrates FcγRIIIa expression



Effector cell line is a Jurkat cell line engineered to express FcγRIIIa (V158) receptor and NFAT-RE luc2 luciferase



| | EC50 |
|----------------------|------------|
| no blocking Ab | 7.612e-010 |
| non specific IgG1 | 6.189e-010 |
| CD16 Ab, clone 3G8 | 3.164e-009 |
| CD16 Ab, clone LNK16 | 5.563e-009 |
| CD16 Ab, clone GRM1 | 9.218e-010 |

Assay used WIL2-S target cells

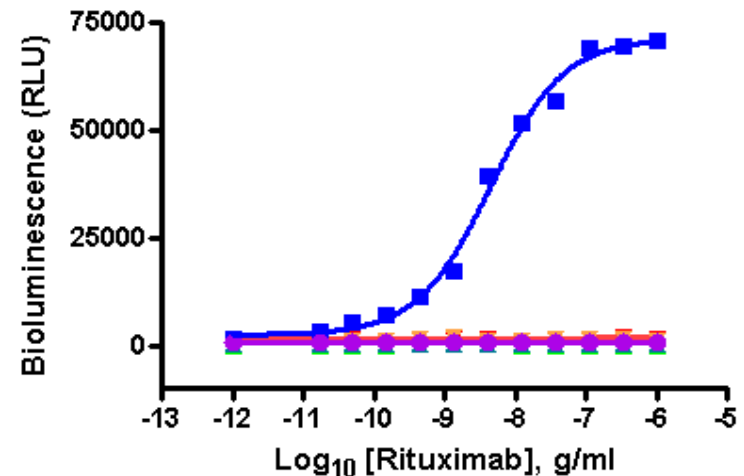
ADCC Reporter Bioassay demonstrates appropriate specificity



- Target cells, effector cells and specific antibody ■ Wil2-S, Jurkat/NFAT-luc+FcγRIIIa, Rituximab
- No Target cells ● NO Wil2-S, Jurkat/NFAT-luc+FcγRIIIa, Rituximab
- No Effector cells or no FcγRIIIa { ▲ Wil2-S, Jurkat/NFAT-luc (NO FcγRIIIa), Rituximab
▲ Wil2-S, NO Jurkat/NFAT-luc+FcγRIIIa, Rituximab
- No antibody or non-specific antibody { ▼ Wil2-S, Jurkat/NFAT-luc+FcγRIIIa, NO Rituximab
▼ Wil2-S, Jurkat/NFAT-luc+FcγRIIIa, Trastuzumab

Assay signal is dependent on:

Presence of Target cells
+
Presence of FcγRIIIa receptor
+
Appropriate specific antibody

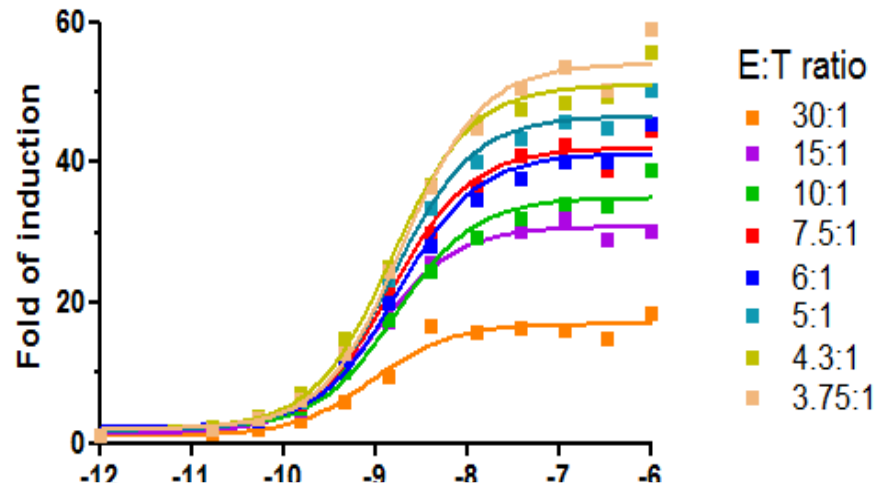


Bioassay development: Optimizing Effector:Target (E:T) ratio



An example: Evaluation of different E:T ratios for assay optimization using frozen, thaw-and-use engineered Jurkat Effector cells (FcγRIIIA (V158) / NFAT-RE-luc2) with fresh-from-culture target cells (WIL2-S, CD20+).

- Fixed Effector cell number
- Varied Target cell number and E:T ratios
- 6hr induction
- Bio-Glo™ Assay Reagent



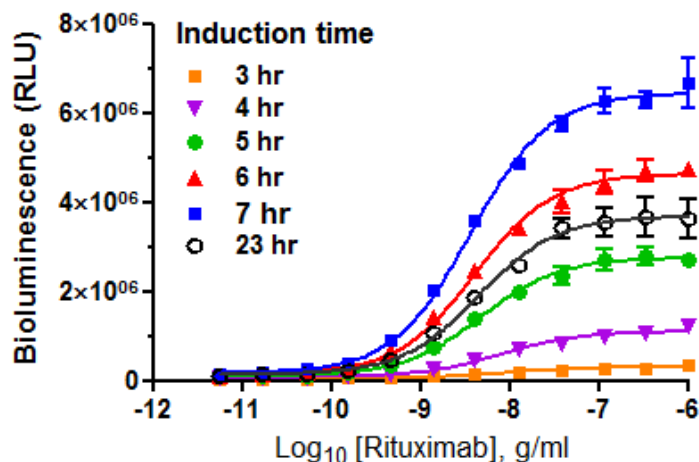
Higher target cell numbers generate greater fold induction

Bioassay development: Evaluating bioassay duration for response (fold induction)

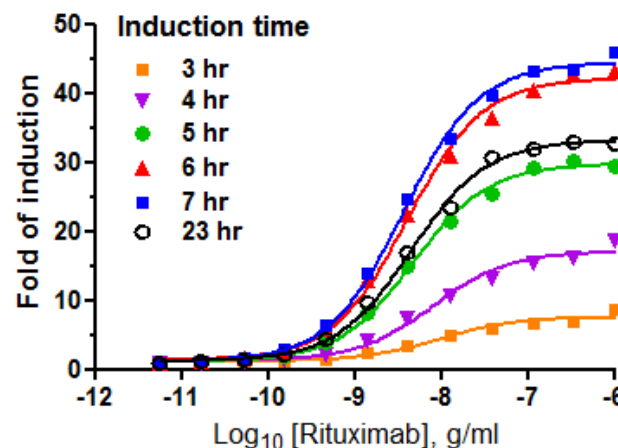


Evaluation of different induction hours for assay optimization using frozen, thaw-and-use Jurkat Effector cells with fresh-from-culture target cells (WIL2-S, CD20+)

Bioluminescence



Fold Induction



| | 3 hr | 4 hr | 5 hr | 6 hr | 7 hr | 23 hr |
|------|------------|------------|------------|------------|------------|------------|
| EC50 | 8.978e-009 | 7.755e-009 | 4.605e-009 | 3.968e-009 | 3.577e-009 | 4.352e-009 |

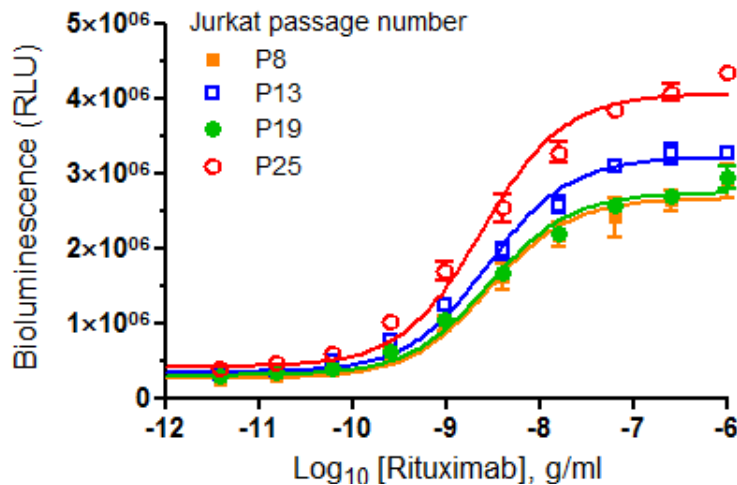
Bioassay can be conducted with as few as 5-6hr induction using frozen, thaw-and-use Effector cells.

Bioassay development: Evaluating clone stability for engineered Jurkat effector cells

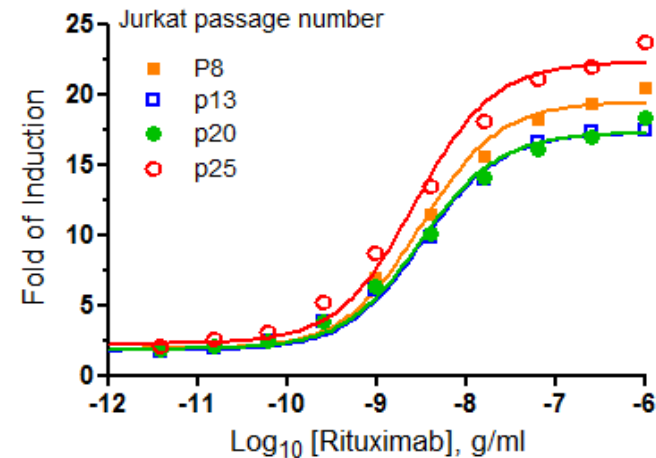


Evaluation of stable effector cell line clone stability for assay optimization using frozen, thaw-and-use effector cells with fresh-from-culture WIL2-S target cells.

Bioluminescence



Fold Induction



| | P8 | p13 | p20 | p25 |
|------|------------|------------|------------|------------|
| EC50 | 3.094e-009 | 3.211e-009 | 3.077e-009 | 2.665e-009 |

*Frozen, thaw-and-use effector cell responsiveness was stable up to passage 25.
(Current stability > passage 50)*

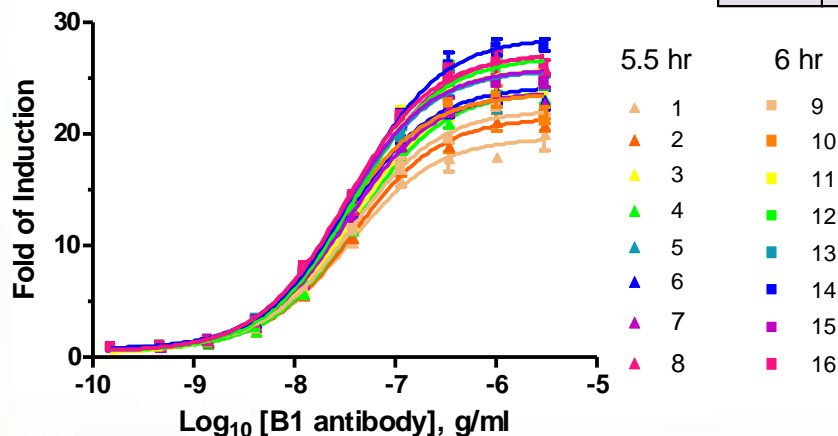
- E:T ratio = 7.5:1
- 6hr induction
- Bio-Glo™ Assay Reagent

Bioassay development: Optimization using DOE

Variables:

1. Induction time
2. Target/Ab pre-incubation
3. Effector cell number
4. Target cell number

| run | induction time hr | Target cell / Ab | |
|-----|-------------------|-----------------------|--------------------|
| | | incubation time(mins) | plating number (K) |
| 1 | 5.5 | 30 | 75 |
| 2 | 5.5 | 30 | 75 |
| 3 | 5.5 | 30 | 90 |
| 4 | 5.5 | 30 | 90 |
| 5 | 5.5 | 45 | 75 |
| 6 | 5.5 | 45 | 75 |
| 7 | 5.5 | 45 | 90 |
| 8 | 5.5 | 45 | 90 |
| 9 | 6 | 30 | 75 |
| 10 | 6 | 30 | 75 |
| 11 | 6 | 30 | 90 |
| 12 | 6 | 30 | 90 |
| 13 | 6 | 45 | 75 |
| 14 | 6 | 45 | 75 |
| 15 | 6 | 45 | 90 |
| 16 | 6 | 45 | 90 |



Outputs and Results:

Good response (fold induction) = 19-27

Good (low) L-term values = 0.1-0.2*

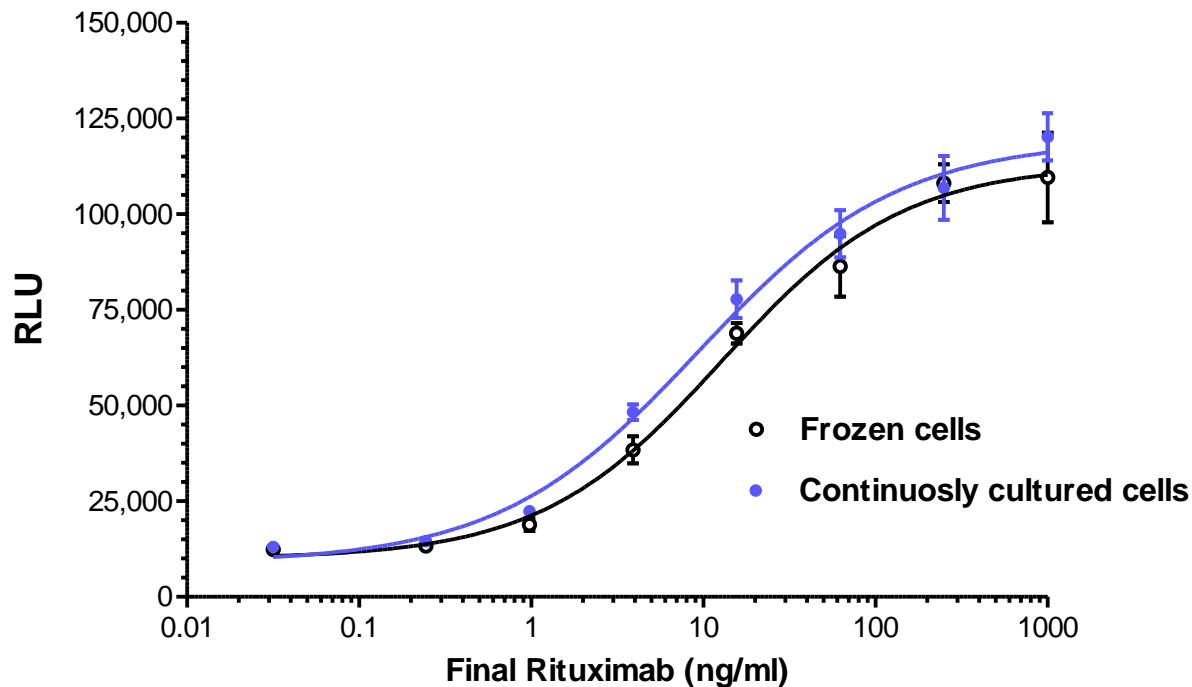
* a measure of assay precision around the EC₅₀ determination
(log width of the 95% confidence interval around logEC₅₀)

Target cells can also be frozen, thaw-and-use format



WIL2-S target cells can be used in frozen, thaw-and-use format

Direct comparison of target cell format: continuously-cultured vs. frozen, thaw-and-use cells



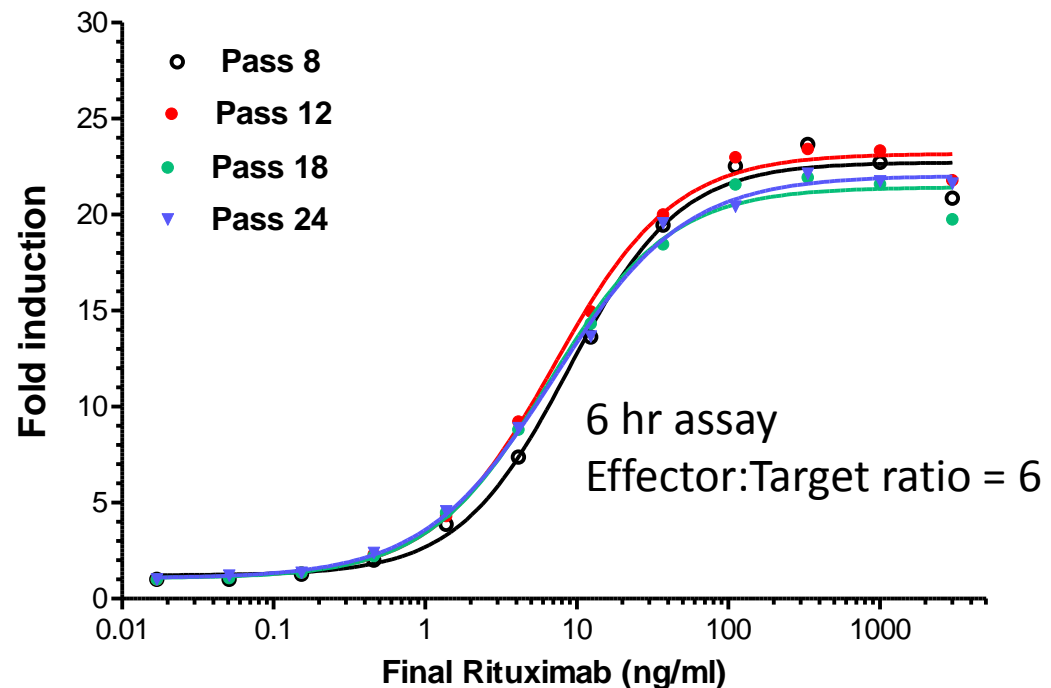
Effector cells: FcγRIIIa/NFAT-luc Jurkat (frozen, thaw-and use); Ab: Rituximab

Evaluating passage and cell density at freezing for WIL2-S (CD20+) Target cells



Establishment of optimized frozen, thaw-and-use Target cells includes establishment of acceptable passage range and cell growth conditions prior to freezing.

Important for establishing cell banks and manufacturing capabilities

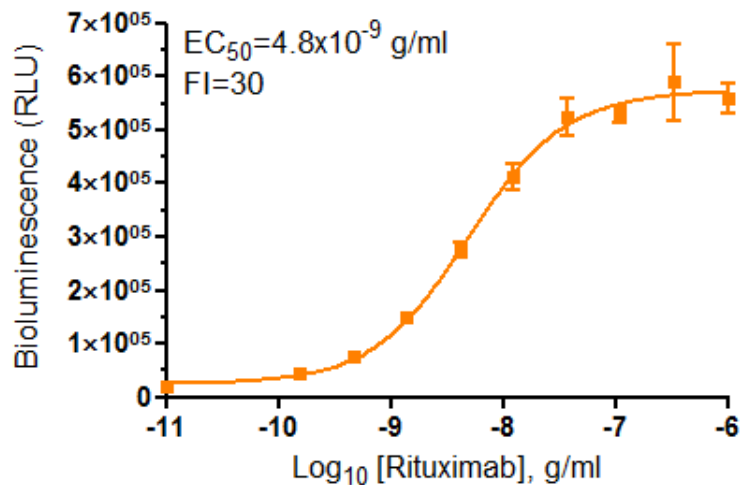


| | Pass 8 | Pass 12 | Pass 18 | Pass 24 |
|------|--------|---------|---------|---------|
| EC50 | 8.851 | 7.057 | 6.520 | 7.122 |

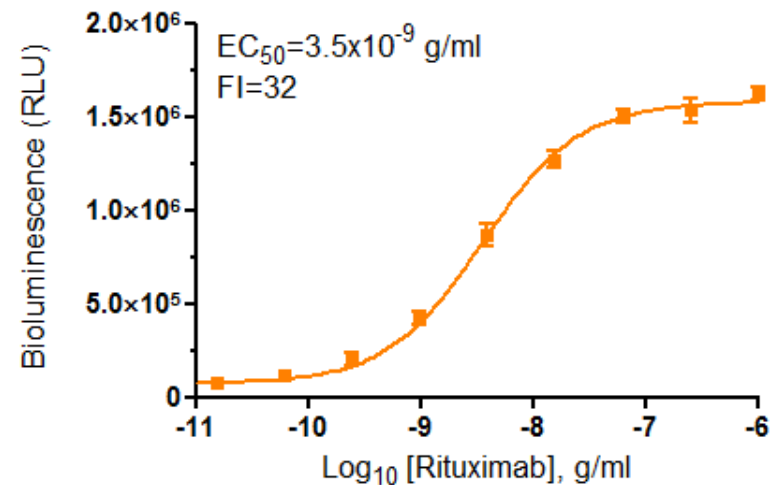
ADCC Reporter Bioassay using frozen, thaw-and-use Effector and Target cells



A. Fresh cells from continuous culture



B. Frozen, thaw-and-use cells



Target cells are CD20+ WIL2-S

EC₅₀s and fold induction of responses are equivalent

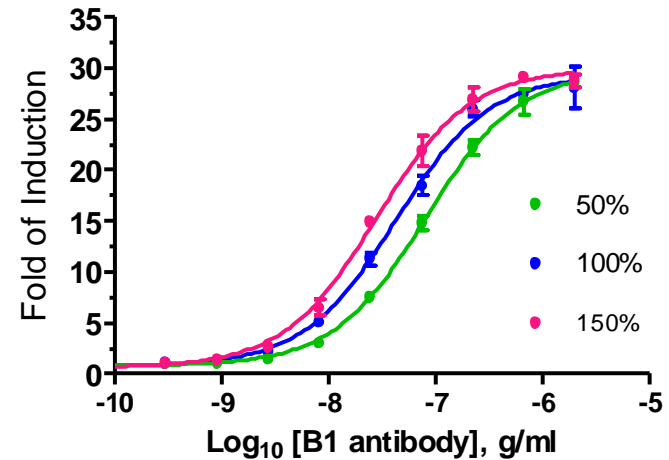
Performance characteristics of ADCC Reporter Bioassay



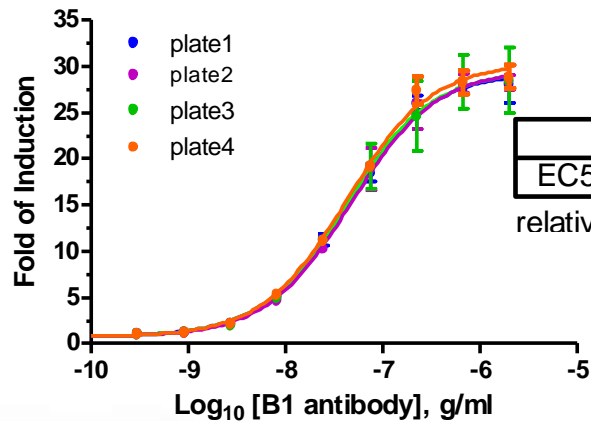
Design:

- Two analysts
- Three days
- Four plates per day
- ✓ 100% vs 50%
- ✓ 100% vs 75%
- ✓ 100% vs 125%
- ✓ 100% vs 150%

Measure relative potency and parallelism



Measure repeatability



| | 100%-1 | 100%-2 | 100%-3 | 100%-4 |
|------------------|------------|------------|--------------|------------|
| EC50 | 4.230e-008 | 4.693e-008 | 4.323e-008 | 4.245e-008 |
| relative potency | 102% | 92% | 100% control | 102% |

Assay with frozen, thaw-and-use effector and WIL2-S target cells

Precision, accuracy and linearity of ADCC Reporter Bioassay



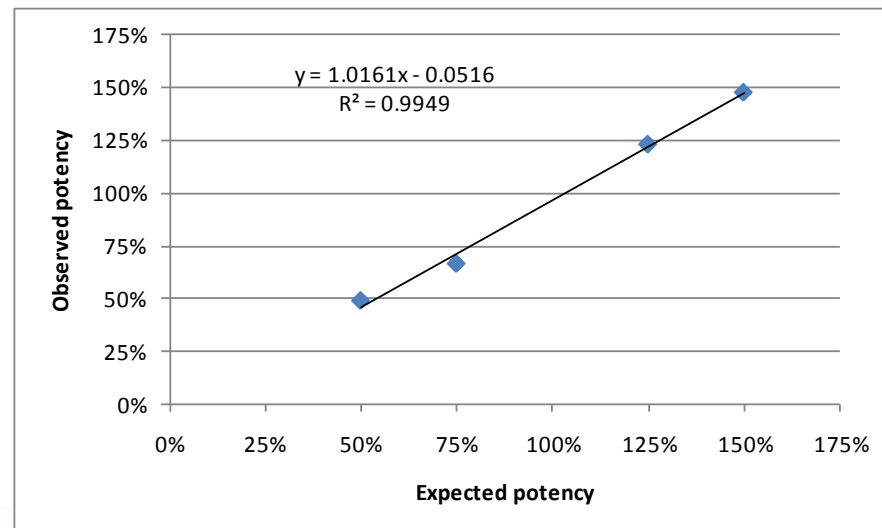
| | Antibody Sample | Measured Potency (%) | Mean Potency (%) | SD % | Recovery (%) | RSD (%) |
|-------|-----------------|----------------------|------------------|------|--------------|---------|
| day 1 | | 48.5 | | | | |
| day 2 | 50% | 45.2 | 48.9 | 3.9 | 97.7 | 7.9 |
| day 3 | | 52.9 | | | | |
| day 1 | | 63.1 | | | | |
| day 2 | 75% | 62.9 | 66.4 | 5.9 | 88.5 | 8.9 |
| day 3 | | 73.2 | | | | |
| day 1 | | 112.1 | | | | |
| day 2 | 125% | 136.3 | 123.0 | 12.3 | 98.4 | 10.0 |
| day 3 | | 120.5 | | | | |
| day 1 | | 148.8 | | | | |
| day 2 | 150% | 150.4 | 147.6 | 3.6 | 98.4 | 2.4 |
| day 3 | | 143.6 | | | | |

Precision
(Av of RSD(%))
7.3%

Accuracy
(Av of % Recovery)
95.8%

Linearity
 $y = 1.016x - 0.052$
 $R^2 = 0.995$

Assay with frozen, thaw-and-use effector and WIL2-S target cells



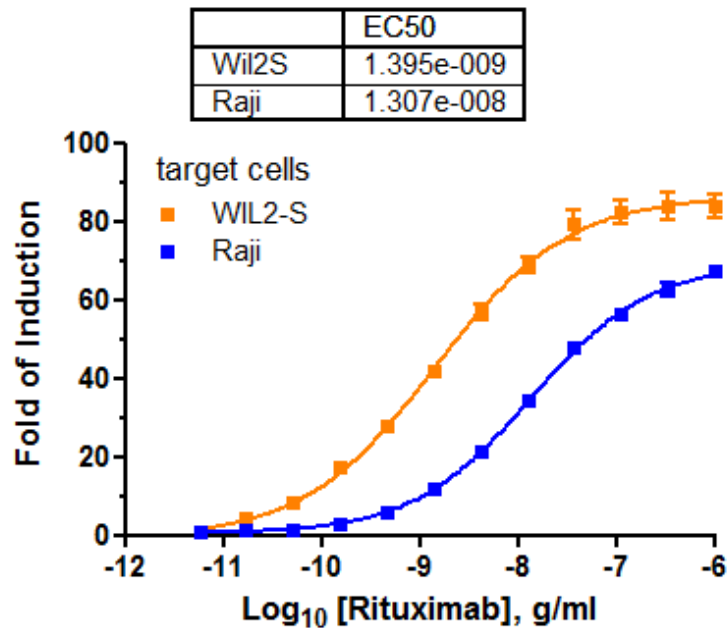
ADCC Reporter Bioassay can measure potency of antibody drugs in different systems



Suspension or adherent target cells can be used

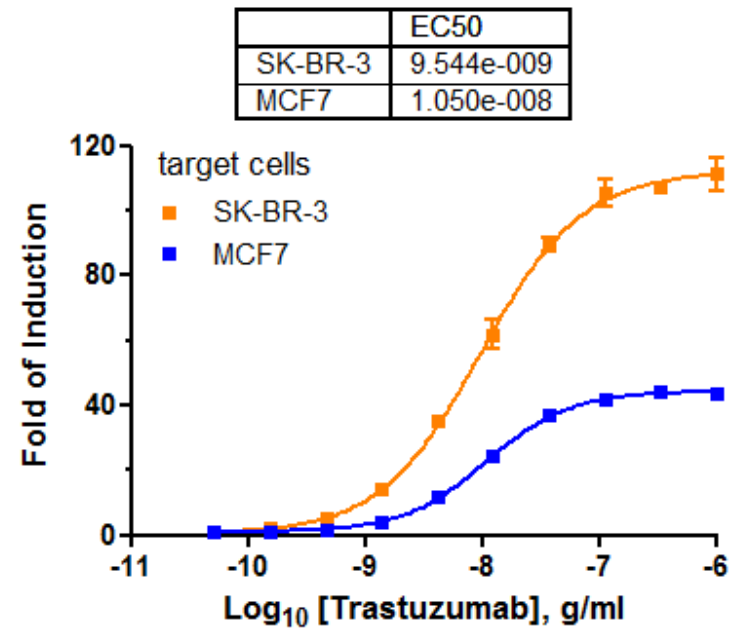
Rituximab (anti-CD20)

CD20⁺ B cell lines (suspension) as target cells
(plated directly from suspension)



Trastuzumab (anti-Her-2)

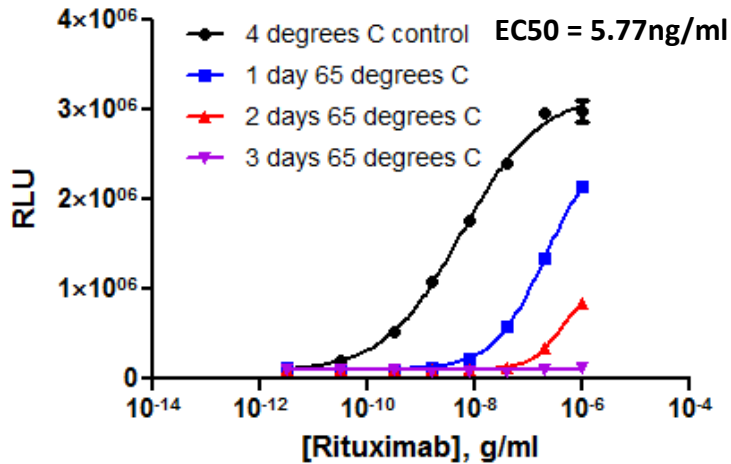
Her2⁺ breast cancer cell lines (adherent)
as target cells (plated 16-18hr prior)



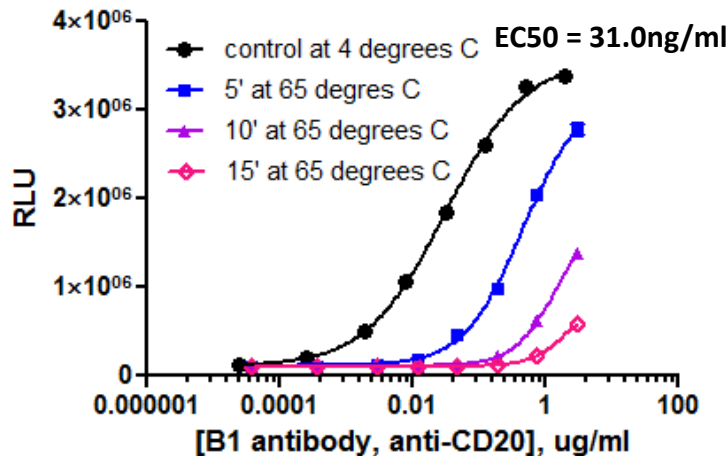
ADCC Reporter Bioassay is stability-indicating for Fc effector function



Rituximab



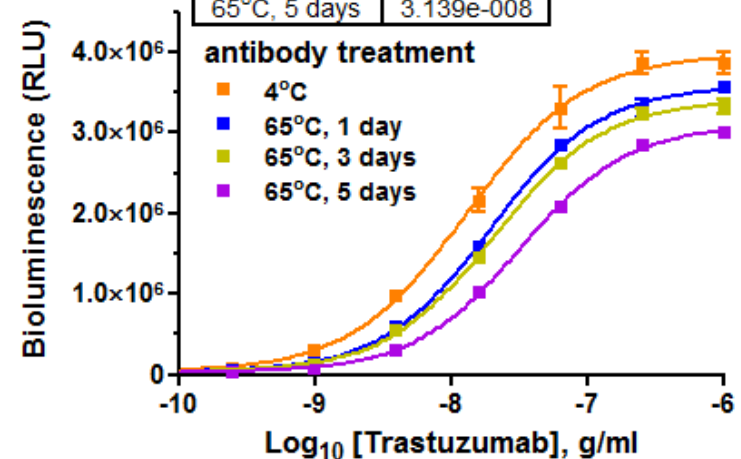
Tositumomab



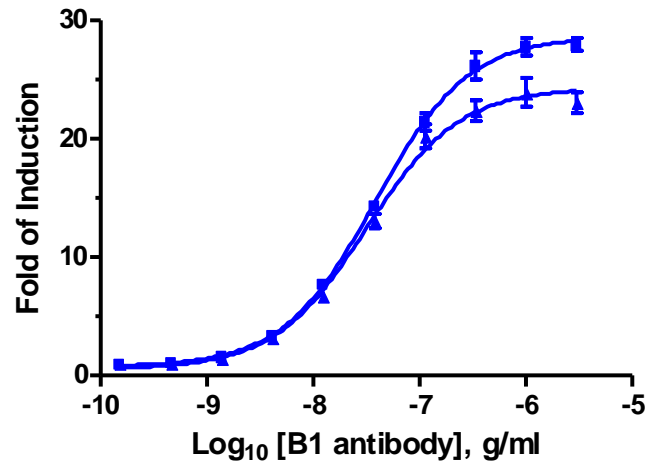
Activity of heat-treated antibody drugs

Trastuzumab

| | EC50 |
|--------------|------------|
| 4°C | 1.284e-008 |
| 65°C, 1 day | 1.902e-008 |
| 65°C, 3 days | 2.031e-008 |
| 65°C, 5 days | 3.139e-008 |

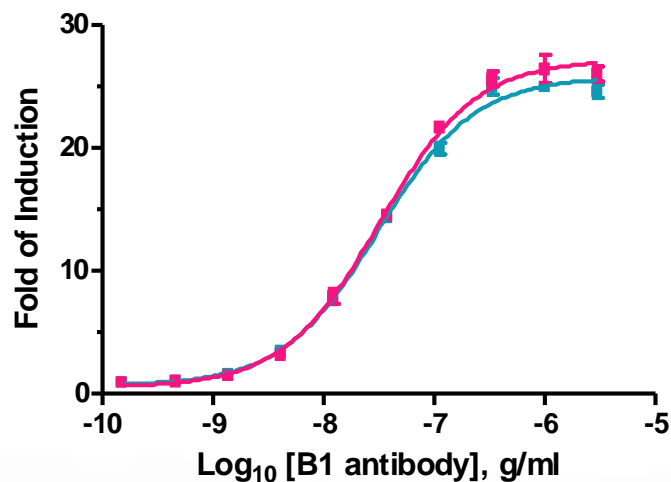


ADCC Reporter Bioassay is robust



Time of induction

| Run | Hr induction | EC50 |
|-----|--------------|----------------------------|
| 1 | 6 hr | 3.15×10^{-8} g/ml |
| 2 | 5.5 hr | 3.83×10^{-8} g/ml |

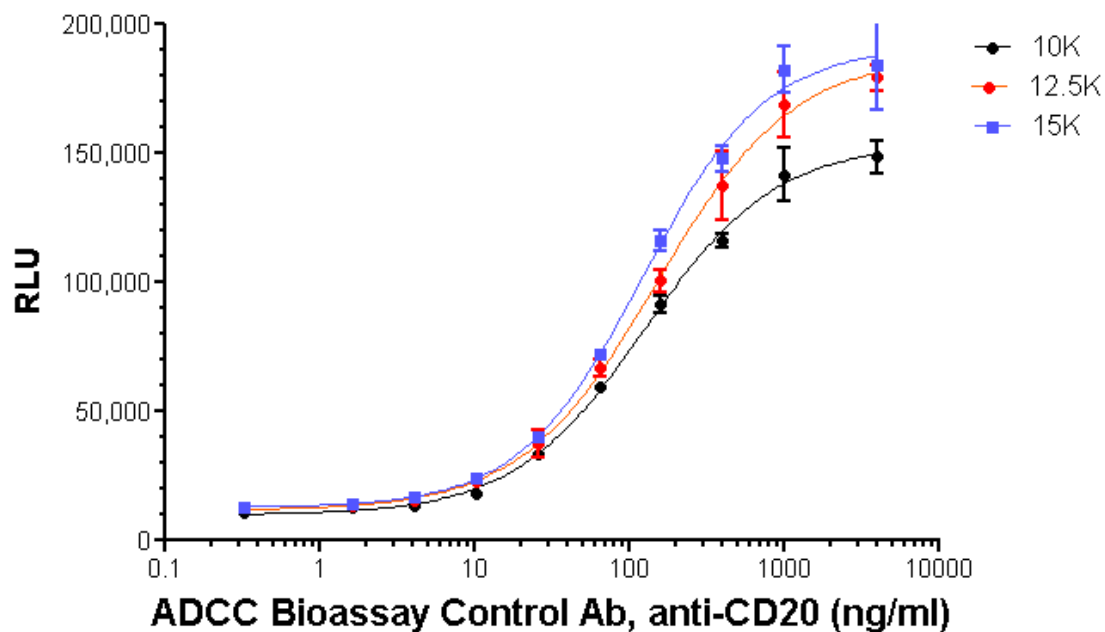


E:T ratio and cells/well

| Run | E:T ratio | T cells | E cells | EC50 |
|-----|-----------|---------|---------|----------------------------|
| 1 | 7.5:1 | 10K | 75K | 3.09×10^{-8} g/ml |
| 2 | 6:1 | 15K | 90K | 3.83×10^{-8} g/ml |

Additional target cell option: Raji cells

Frozen, thaw-and-use FcγRIIIa/NFAT-luc2 Effector Cells (75K per well)



| | 10K | 12.5K | 15K |
|------|-------|-------|-------|
| EC50 | 125.5 | 150.3 | 122.3 |

Accuracy, linearity & precision using frozen, thaw-and-use Raji cells and CD20 Ab



Analyst 1

| Day | Antibody Sample | Measured Potency (%) | Mean Potency (%) | SD (%) | Recovery (%) | CV (%) |
|-----|-----------------|----------------------|------------------|--------|--------------|--------|
| 1 | 50% | 49.9 | 51 | 0.7 | 102 | 1.4 |
| 2 | | 51.3 | | | | |
| 3 | | 50.5 | | | | |
| 1 | 75% | 78.9 | 76 | 5.11 | 101 | 6.7 |
| 2 | | 78.8 | | | | |
| 3 | | 70 | | | | |
| 1 | 125% | 118.6 | 117 | 1.19 | 94 | 1 |
| 2 | | 116.9 | | | | |
| 3 | | 116.3 | | | | |
| 1 | 150% | 143.2 | 145 | 3.91 | 97 | 2.7 |
| 2 | | 142.5 | | | | |
| 3 | | 149.6 | | | | |

Precision: 2.95%

Accuracy: (recovery average): 98.5%

**Linearity:
Y=0.922x+5.0**

Analyst 2

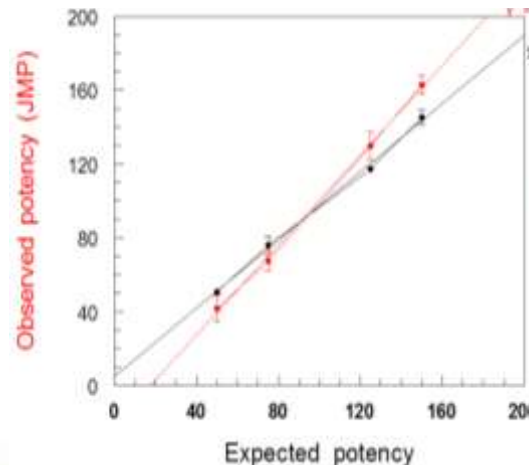
| Day | Antibody Sample | Measured Potency (%) | Mean Potency (%) | SD (%) | Recovery (%) | CV (%) |
|-----|-----------------|----------------------|------------------|--------|--------------|--------|
| 1 | 50% | 38.4 | 41.8 | 7.2 | 83.5 | 17.2 |
| 2 | | 47.2 | | | | |
| 3 | | 33.2 | | | | |
| 4 | | 48.2 | | | | |
| 1 | 75% | 59.6 | 67.4 | 5.2 | 89.9 | 7.7 |
| 2 | | 70.2 | | | | |
| 3 | | 69.3 | | | | |
| 4 | | 70.5 | | | | |
| 1 | 125% | 120 | 129.7 | 7.5 | 103.7 | 5.8 |
| 2 | | 132.3 | | | | |
| 3 | | 137.8 | | | | |
| 4 | | 128.6 | | | | |
| 1 | 150% | 160.2 | 162.8 | 5.2 | 108.5 | 3.2 |
| 2 | | 158.2 | | | | |
| 3 | | 162.7 | | | | |
| 4 | | 170 | | | | |

Precision: 8.47%

Accuracy (recovery average): 96.4%

**Linearity:
Y=1.22x-21.3**

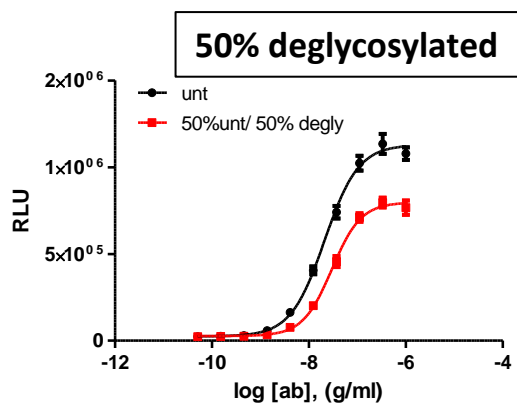
Linearity



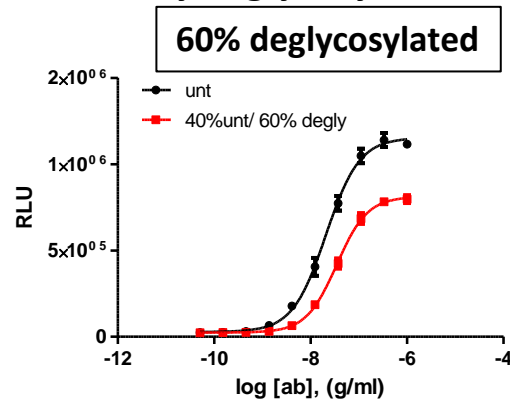
Detection of small differences in Fc effector biological activity of therapeutic antibody preps



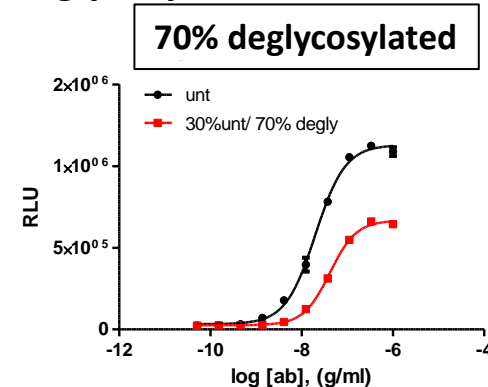
Differentiation of blended % mixes of fully N-glycosylated & fully deglycosylated Trastuzumab



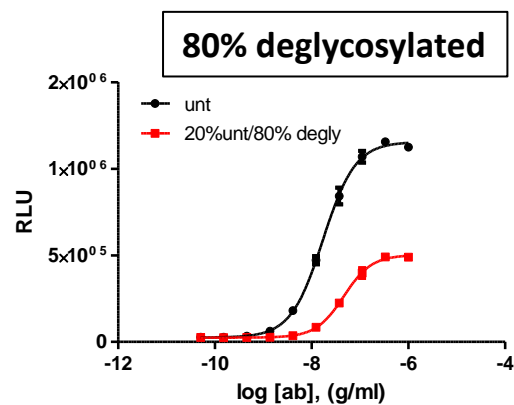
| | unt | 50%unt/ 50% degly |
|------|------------|-------------------|
| EC50 | 2.110e-008 | 2.990e-008 |



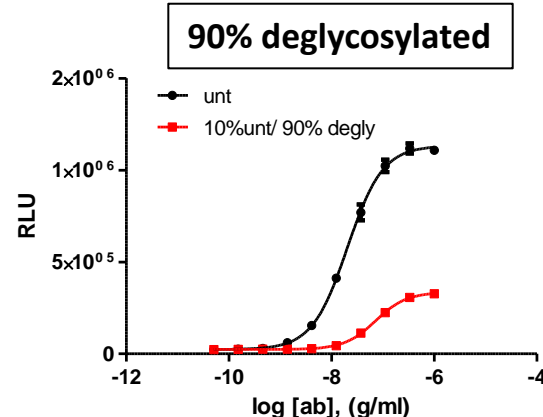
| | unt | 40%unt/ 60% degly |
|------|------------|-------------------|
| EC50 | 2.082e-008 | 3.486e-008 |



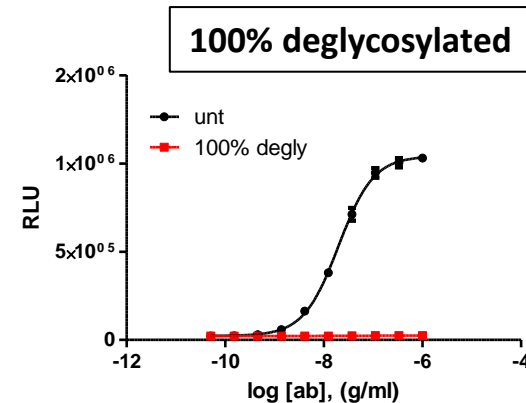
| | unt | 30%unt/ 70% degly |
|------|------------|-------------------|
| EC50 | 2.002e-008 | 4.153e-008 |



| | unt | 20%unt/80% degly |
|------|------------|------------------|
| EC50 | 1.720e-008 | 4.626e-008 |



| | unt | 10%unt/ 90% degly |
|------|------------|-------------------|
| EC50 | 2.037e-008 | 7.174e-008 |



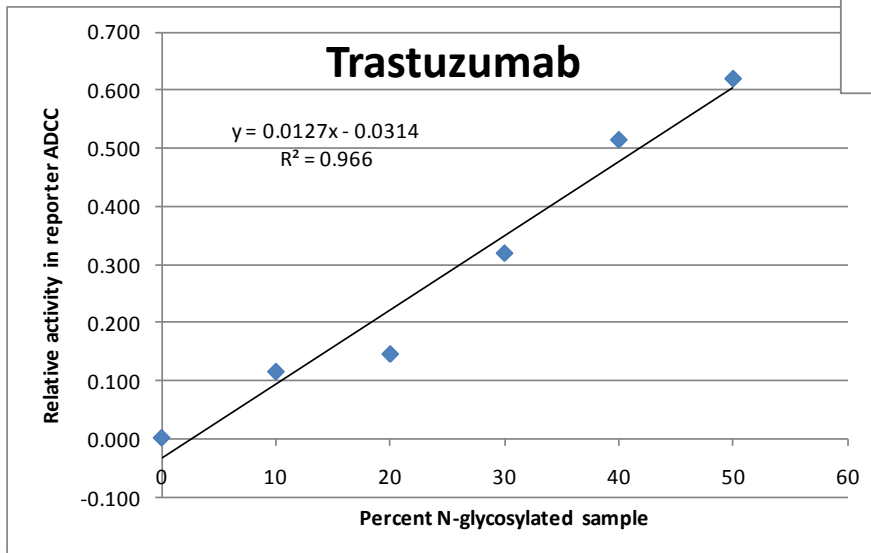
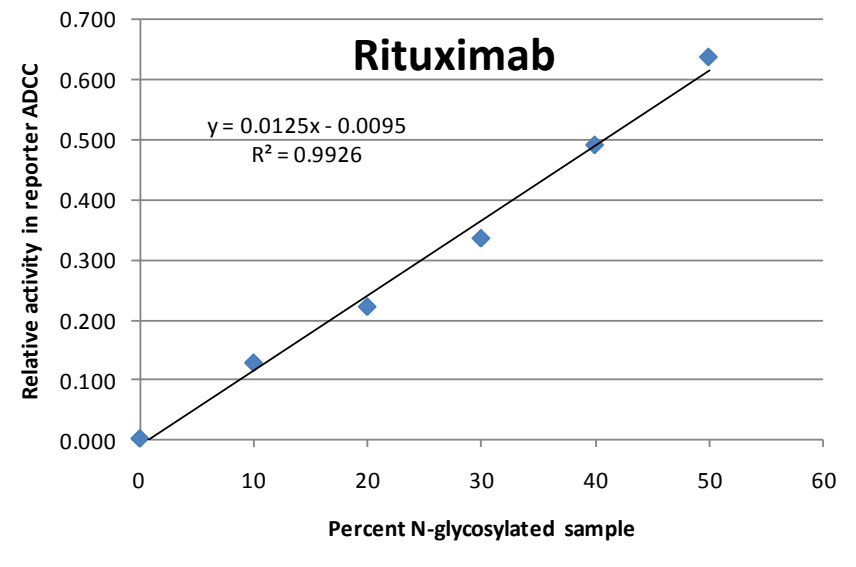
| | unt | 100% degly |
|------|------------|------------|
| EC50 | 1.988e-008 | 3.202e-008 |

Target cells: SKBR3; Unt = 100% glycosylated

Fc effector biological activity correlates well with antibody N-glycosylation



Rituximab and Trastuzumab:
Good correlation between blended % mixes of fully N-glycosylated and deglycosylated preparations and their relative biological activity in ADCC Reporter Bioassay



Small differences in Fc effector activity in ADCC MOA pathway are easily distinguished in the ADCC reporter bioassay

Planned ADCC Reporter Bioassay kit configurations



Core Kit

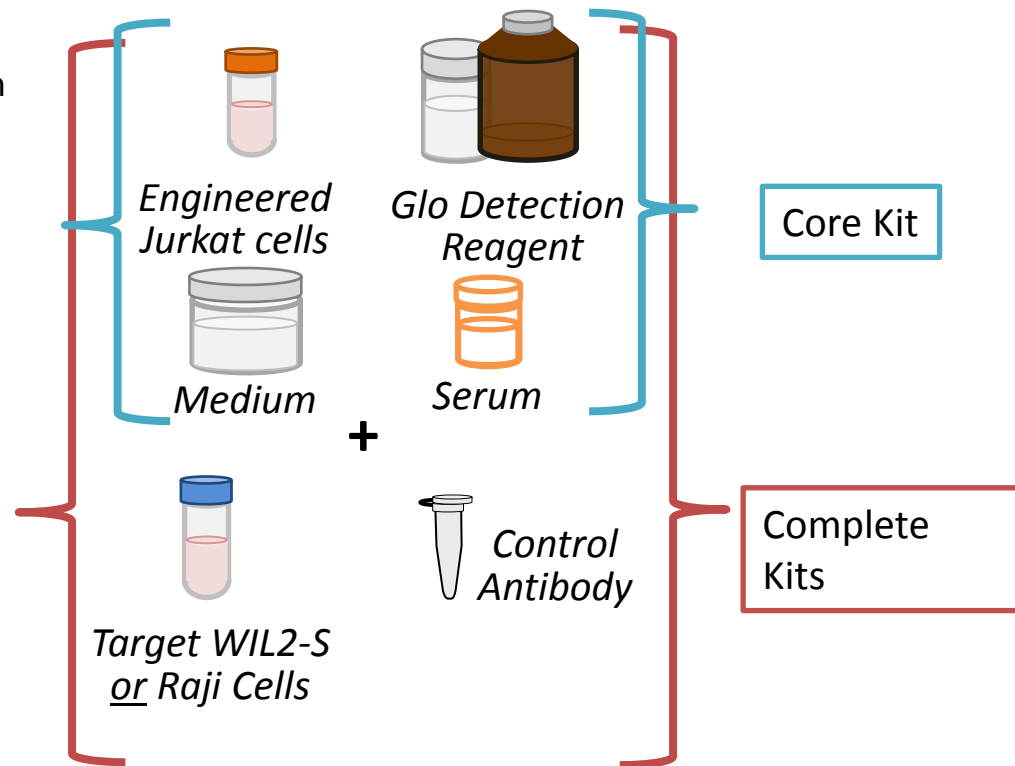
- Engineered Jurkat Effector cells (NFAT-RE-luc2 / FcγRIIIa); *frozen, thaw-and-use*
- Bio-Glo™ Luciferase Assay System
- RPMI Medium
- Low IgG Serum

Use with different Abs and target cells

Complete Kits (2 types)

- *Above PLUS:*
- Target WIL2-S cells; *frozen, thaw-and-use* or Target Raji cells; *frozen thaw-and-use*
- Control Ab (CD20)

Use as assay control, or when WIL2-S or Raji target cells are suitable or target is CD20



Summary of the ADCC Reporter Bioassay

Design Features

1. Low variability
2. Engineered effector cells to replace primary NK cells (Jurkat FcγRIIIa/NFAT-RE-luc2)
3. “Cells as reagents”, frozen, thaw-and-use format – consistency & convenience
4. Simplicity and robustness of assay protocol and reagents
5. Broad applicability to use with multiple target cells – suspension or adherent

Benefits

- ✓ Demonstrates precision, accuracy, linearity, robustness
- ✓ Can quantify potency and stability of therapeutic Ab drugs
- ✓ Can differentiate differences in relative biological activity of Fc effector function in ADCC MOA that result from small changes in Ab glycosylation

For more information



Neal Cosby, PhD
Strategic Marketing Manager
Neal.cosby@promega.com

Or

Custom Order Department
COD@promega.com