

microRNA Purification from Plant Tissues Using the ReliaPrep™ miRNA Cell and Tissue Miniprep System

Purify high-quality, amplifiable total RNA, including microRNA, from plant tissue.

Promega Corporation

Sample Type: Corn, soybean and *Arabidopsis* leaf tissue.

Material Required: Fisherbrand® Motorized Tissue Grinder (Fisher Scientific Cat.# 12-1413-61).

Analyses: UV absorbance and QuantiFluor® quantitation, RT-qPCR

Protocol: ReliaPrep™ miRNA Cell and Tissue Miniprep System Technical Manual #TM469.

Disclaimers:

This protocol was developed by Promega Applications Scientists and is intended for research use only.

Users are responsible for determining suitability of the protocol for their application.

Further information can be found in Technical Manual #TM469, available at: www.promega.com/protocols, or e-mail: techserv@promega.com

Protocol

1. Grind plant tissue using liquid nitrogen and mortar and pestle.
2. Add 200µl of LBA + 1-thioglycerol to plant tissue in a 1.5ml microcentrifuge tube.
Note: Additional LBA + 1-thioglycerol may be added to maximize recoverable lysate volume.
3. Homogenize the tissue using a Fisherbrand® Motorized Tissue Grinder with disposable pestles.
4. Centrifuge the sample at maximum speed for 1 minute to pellet debris. Transfer the cleared homogenate (maximum 200µl) to a clean 1.5ml microcentrifuge tube.
5. Add 130µl of RDB, and vortex for 10 seconds.
6. Centrifuge at 12,000 × *g* for 2 minutes.
7. Transfer homogenate to a new 1.5ml tube.
8. Add 400µl of 100% isopropanol to each cleared homogenate. Mix by vortexing.
9. Transfer homogenate to a ReliaPrep™ Minicolumn. Centrifuge at 12,000 × *g* for 30 seconds.
10. Discard the liquid in the collection tube.
11. Add 500µl of RWA to each column. Centrifuge at 12,000 × *g* for 30 seconds. Discard liquid in the collection tube.
12. Add 500µl of RWA to each column. Centrifuge at 12,000 × *g* for 2 minutes.
13. Transfer column to a 1.5ml Elution Tube.
14. Add 40µl of Nuclease-Free Water to each column. Centrifuge at 12,000 × *g* for 1 minute.
15. Transfer 5µl of DNase I and 5µl of DNase 10X Buffer to each eluate.
16. Incubate 5 minutes at room temperature.
17. Add 150µl of LBA to the samples.

18. Add 300µl of 95% ethanol to the mixture, and vortex for 10 seconds. Transfer mixture to a new ReliaPrep™ Minicolumn.
19. Centrifuge at 12,000 × *g* for 30 seconds. Discard the liquid in the collection tube.
20. Add 500µl of RWA. Centrifuge at 12,000 × *g* for 30 seconds. Discard liquid in the collection tube.
21. Add 500µl of RWA. Centrifuge at 12,000 × *g* for 2 minutes. Discard liquid in the collection tube.
22. Transfer column to a 1.5ml Elution Tube.
23. Add 50µl of Nuclease-Free Water. Centrifuge at 12,000 × *g* for 1 minute.

Results

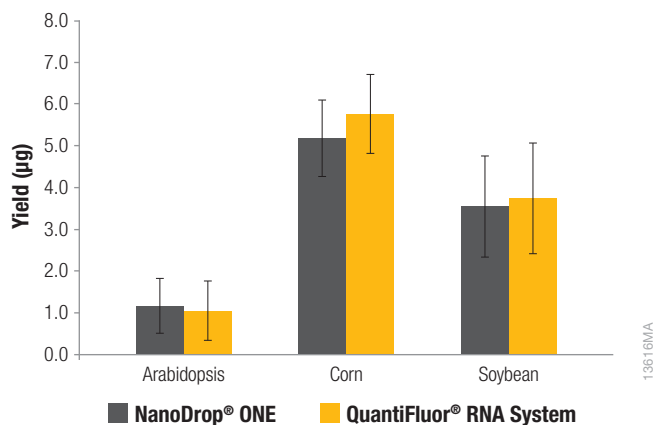


Figure 1. Yields of RNA from plant tissues determined using the NanoDrop® ONE and the QuantiFluor® RNA System (Cat.# E3310) on the Quantus™ Fluorometer (Cat.# E6150). N=3.

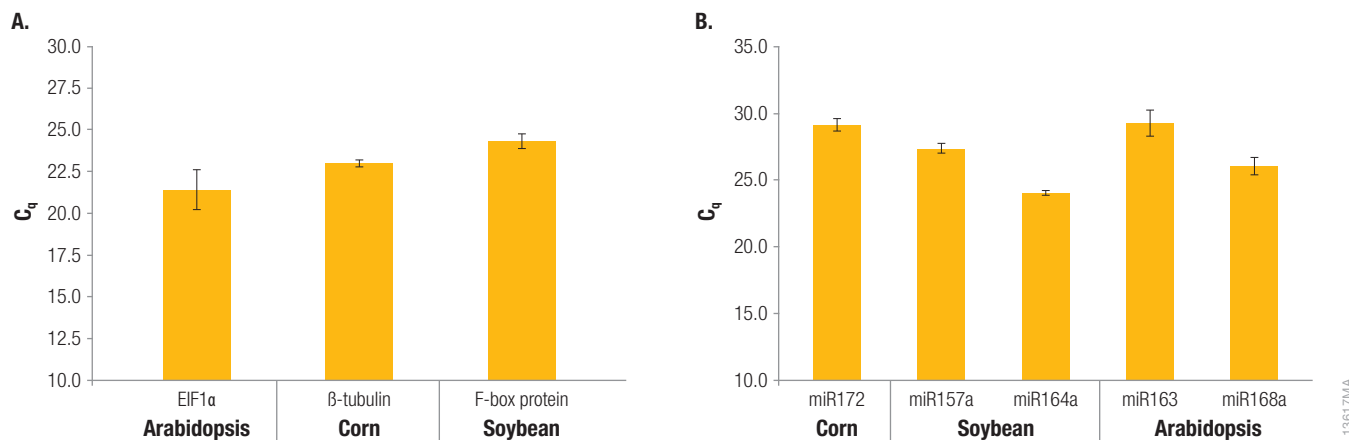


Figure 2. RT-qPCR of purified RNA. Panel A. RT-qPCR of mRNA using plant-specific primers for housekeeping genes with the GoTaq® 1-Step RT-qPCR System (Cat.# A6020). N=3. Panel B. RT-qPCR of miRNA using TaqMan® miRNA-specific primers and the TaqMan® microRNA reverse transcription kit followed by qPCR using the GoTaq® Probe qPCR Master Mix (Cat.# A6101). N=3.

Ordering Information

Product

ReliaPrep™ miRNA Cell and Tissue Miniprep System

Size	Cat.#
10 preps	Z6210
50 preps	Z6211
250 preps	Z6212

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